



resDetect™ *E. coli* resDNA Quantitative Kit (qPCR)

Catalog Number: OPA-O002

Assay Tests: 100 Tests

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures.

IMPORTANT: Please carefully read this user guide before performing your experiment.

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Product Information






The *E. coli* resDNA Quantitative Kit is designed for quantitative detection of residual *E. coli* DNA in biopharmaceutical productions (Antibodies, Cells, Vaccines, etc.). Use the kit after extracting host cell DNA from test samples. To achieve better DNA recovery, it is recommended to use the **resDetect™ resDNA Sample Preparation Kit (Magnetic Beads) (Cat. No. OPA-R005)** in combination with this kit. For more information about the extraction kit, refer to the resDNA Sample Preparation Kit User Guide (ACROBiosystems.com).

Residual *E. coli* DNA is quantified using a real-time polymerase chain reaction (PCR) assay. The PCR-based assay is sensitive and specific for DNA from *E. coli* genome and not subject to detection of human or environmental DNA that might be introduced during sample handling. The kit was developed to meet the sensitivity requirements defined by WHO (10 ng *E. coli* DNA per therapeutic dose).

To generate the standard curve used to quantitate the DNA in test samples, the *E. coli* assays require six dilutions. The *E. coli* Control DNA for standard curve generation is included in the kits. Linearity is demonstrated by analysis of standard DNA from *E. coli* ranging from 3 fg/μL~300 pg/μL. Limit of Detection is 1 fg/μL.

Contents and Storage

The kit contains sufficient reagents to run 100 PCR reactions each with a final reaction volume of 30 μ L.

Contents	Colors	Amount	Storage
2 \times qPCR Master Mix		1.0 mL \times 2	-30°C to -15°C Note: 2 \times qPCR Master Mix and Primer & Probe Mix need protection from light.
<i>E. coli</i> Primer & Probe Mix		700 μ L \times 1	
<i>E. coli</i> DNA Control (3 ng/ μ L)		100 μ L \times 1	
Dilution Buffer		1.5 mL \times 3	
DNase/RNase-free Water		1.0 mL \times 1	

The unopened kit can be stored stably for 18 months from the date of manufacture when kept at a storage temperature between -30°C and -15°C.



Required materials not supplied


Equipment	Real-time PCR instrumentation
Consumables	96-Well Reaction Plate, Covers
	Nuclease-free, DNA-free aerosol-resistant pipet tips
	Low DNA-Binding Microcentrifuge Tubes (Nuclease-free, DNA-free) to prepare working solution, dilutions and mixes.

Workflow

Step 1 Prepare the kit reagent and standard curve dilution


- 2 × qPCR Master Mix
- Primer & Probe Mix
- DNA Control
- Dilution Buffer
- DNase/RNase-Free Water

 for 20~30s
 Briefly centrifuge



Preparation of ERC: Taking a spiking amount of 3 pg as an example, add 10 μL of SD4 (300 fg/μL) to 100 μL of the sample to be tested, vortex it thoroughly, and this is the ERC.
Note: The SD4 used for spiking in the ERC should be from the same tube as the SD4 in the standard curve samples.

Step 2 Prepare the PCR reaction mix

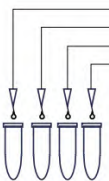


Working Mix (for 1 reaction)
 +15 μL 2 × qPCR Master Mix (uncolored cap)
 +5 μL Primer & Probe Mix (brown cap)




+20 μL Working mix

Step 3 Add samples and controls


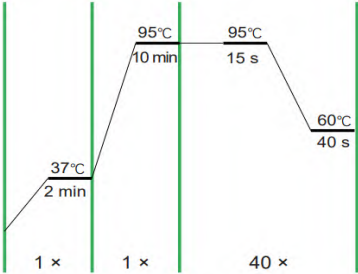


Sample (+10 μL Sample purified solution)
 ERC (+10 μL ERC purified solution)
 NEC (+10 μL NEC purified solution)
 NTC (+10 μL DNase/RNase-Free Water (green cap))

Close lid tightly.

 for 30 s

Step 4 Create the plate document and run qPCR

+ add
 vortex
 centrifuge

Protocol

Prepare the DNA control serial dilutions for the standard curve

1. Guidelines for standard dilutions

- 1.1 Prepare the standard curve and the test samples in different areas of the lab.
- 1.2 Use Low DNA-Binding microcentrifuge tubes and different sets of pipettors for test sample preparation and for standard curve preparation and aliquoting to avoid cross-contamination of test samples.
- 1.3 Vortex each tube for **20-30 seconds** to ensure thorough mixing of the contents before proceeding with each dilution step.
- 1.4 Briefly centrifuge to collect all the liquid at the bottom before making the next dilution.

2. Prepare the control serial dilutions

- 2.1 Label low DNA-binding microfuge tubes: **SD1, SD2, SD3, SD4, SD5, SD6, NTC**, where **SD** indicates serial dilutions and **NTC** indicates the no template control.
- 2.2 Add **30-40 µL** of DNase/RNase-free Water to tube **NTC**. Put aside.
- 2.3 Add **90 µL** of Dilution Buffer to tubes **SD1, SD2, SD3, SD4, SD5, SD6**.
- 2.4 Remove the tube of *E. coli* DNA control (3 ng/µL) from the freezer.
- 2.5 After the DNA thaws, vortex it gently for 20-30 seconds, then briefly centrifuge to collect the solution at the bottom.
- 2.6 Perform the serial dilutions:
 - a. Add **10 µL** of the DNA control to the tube that is labeled **SD1**, then vortex thoroughly and briefly centrifuge.
 - b. Transfer **10 µL** of the DNA from tube **SD1** to tube **SD2**, then vortex thoroughly and briefly centrifuge.
 - c. Continue to transfer **10 µL** of DNA from the previous dilution tube to the next dilution tube until you add DNA to tube **SD6**. After each transfer, vortex thoroughly, then centrifuge briefly. Process of dilutions is shown in the following figure.

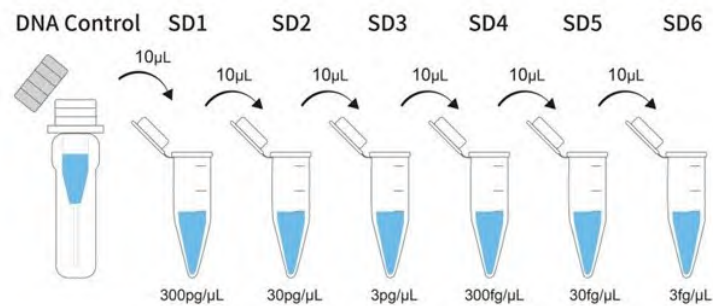


Figure 1. DNA Control Dilution Diagram

2.7 Store the **SD** tubes at 4°C for use within 24 hours.

Preparation and extraction of NEC and ERC (Optional)

1. Preparation and extraction of Negative Extraction Control (NEC)

A Negative Extraction Control (NEC) omits any DNA template from a reaction. This control is used to monitor contamination during nucleic acid extraction. In cases where large numbers of DNA samples need extracted, it is recommended that negative extraction controls are included between the samples for testing.

1.1 Label low DNA-binding 1.5 mL microfuge tubes "NEC".

1.2 Add **100 μL** of 1×PBS (free of Mg²⁺ and Ca²⁺) or 1×TE (pH7.0~pH8.0) to each tube.

When finished, extract DNA from the tubes according to the resDNA Sample Preparation Kit User Guide (Cat. No. OPA-R005), then quantify the extracted DNA in each tube using this Kit (Cat. No. OPA-O002) as described in the following section.

2. Preparation and extraction of Extraction/Recovery Control (ERC)(Optional)

Extraction/Recovery Control (ERC) can be used to evaluate the efficiency of DNA extraction, recovery, and quantitation from test samples. Moreover, the ERC can be used to verify assay and system performance. The following procedure describes the preparation of ERC sample containing *E. coli* DNA control per well for qPCR analysis.

2.1 For each sample, label low DNA binding 1.5 mL microfuge tubes "ERC".

2.2 Add the appropriate volume of test sample to each tube.

2.3 Add **10 µL** of DNA from tube **SD4 (300 fg/µL)** to each ERC tube, then vortex gently.

When finished, extract DNA from the tubes according to the resDNA Sample Preparation Kit User Guide (Cat. No. OPA-R005), then quantify the extracted DNA in each tube using this Kit (Cat. No OPA-O002) as described in the following section.

NOTE: Adjust the amount of *E. coli* DNA control added to the sample for those test samples that contain higher background levels of DNA. To ensure accurate results, the amount of *E. coli* DNA control added to a test sample should be approximately two to three times the amount of DNA measured in the test sample without the addition of the *E. coli* DNA control. To calculate the efficiency of DNA recovery and quantitation from the test samples, subtract the amount of DNA measured in the sample without the addition of *E. coli* DNA control from the amount of DNA measured in the ERC sample.

Prepare the PCR reaction mix

Prepare serial dilutions of *E. coli* DNA control from the same experiment to create a standard curve and to determine sample recovery rate.

1. Determine the number of controls and test samples whose DNA content you will quantify. Number of reaction wells is equal to three times the sum of **NTC, NEC, ERC, SD1-SD6, and test samples**.
2. Thaw reagents completely on ice, thoroughly mix reagent, and briefly centrifuge. Prepare a 2.0 mL tube for **Working Mix** (not add DNA template) using the reagents and volumes shown in the table below, thoroughly mix reagent, and briefly centrifuge.

IMPORTANT! To compensate for pipetting losses, it is recommended that the **N** is equals to number of reaction wells plus 2 or 3.

Kit Reagents	Volume for 1 reaction (30 μ L)	Volume for Working Mix
2 \times qPCR Master Mix	15 μ L	15 μ L \times N
<i>E. coli</i> Primer & Probe Mix	5 μ L	5 μ L \times N
DNA template	10 μ L	Add DNA template to each well separately, not as part of Working mix.
Total	30 μ L	20 μ L \times N

3. Add **20 μ L** Working mix to each well separately.
4. Add **10 μ L** DNA template to the corresponding wells. **Final volume** of PCR reaction is **30 μ L**. It is recommended that the above DNA samples (test samples, NTC, NEC, ERC, and SD) should be placed in different zones during the design and layout of the reaction wells to avoid cross contamination and inaccurate test results.

NOTE: Set up a 96-well PCR plate using the example plate layout shown in the following page.

5. Seal the plate with an optical film, then quick-spin with a centrifuge rotor that is compatible with 96-well plates.

Plate layout

	1	2	3	4	5	6	7	8	9	10	11	12
A									SD1	SD1	SD1	
B	S1	S1	S1		S1(ERC)	S1(ERC)	S1(ERC)		SD2	SD2	SD2	S1
C	S2	S2	S2		S2(ERC)	S2(ERC)	S2(ERC)		SD3	SD3	SD3	S2
D	S3	S3	S3		S3(ERC)	S3(ERC)	S3(ERC)		SD4	SD4	SD4	S3
E									SD5	SD5	SD5	
F									SD6	SD6	SD6	
G	NEC	NEC	NEC									NEC
H									NTC	NTC	NTC	

S=Sample; NTC=No Template Control; NEC=Negative Extraction Control; ERC=Extraction/Recovery Control.

NOTE: The plate layout is a suggested example. Adjust the layout according to the number of test samples to be run.

Create the plate document and run the plate

The following instructions apply only to the ABI 7500 instrument. If you use a different instrument, refer to the applicable instrument guide for setup guidelines.

1. Create a new experiment, and enter the experiment name in the Plate name field.
2. Select the **Quantitation Standard Curve** mode, **TaqMan** reagents, and **Standard** mode.
3. In the Plate Setup, enter the Target Name. Select **FAM** in the Reporter Dye drop-down list. Select **(None)** in the Quencher Dye drop-down list. Select **ROX** in the Passive Reference Dye drop-down list
4. Set up the standard curve as shown in the Plate Layout. Assign the tasks and the enter the appropriate Quantity for each set of triplicates. (SD1-SD6, 300000, 30000, 3000, 300, 30, 3 fg/ μ L).
5. Set up the test samples and controls as shown in the Plate Layout.
6. Set up the qPCR reaction program according to the following Table.
7. Set the reaction volume to **30 μ L**, click "Start Run" in the Run interface to start the qPCR run, and analyze the results after completion.

Step	Temperature	Cycles	Time	Signal Collection
1	37°C	1×	2 min	No
2	95°C	1×	10 min	No
3	95°C	40×	15 s	No
	60°C		40 s	Yes

Analyze the results

After the qPCR run is finished, use the general procedure to analyze the results. The threshold value for ABI7500 is set to 0.2, and use the auto baseline setting. For other instruments, the setting of parameters should be adjusted according to the specific instrument user guide and software version. The acceptance criteria of results are shown as follows:

1. The Standard curve: $R^2 \geq 0.98$, Eff%=90-110%.
2. The detection result of NTC should be undetermined or Ct value > 35.
3. The Ct value of NEC should be greater than the Ct value of the SD6.
4. Ct values should be consistent with replicates, the differences between the Ct values of replicates is less than 0.5.
5. The spike recoveries of test samples and controls should be between 50%-150%. The calculation formula of spiked recovery rate is as follows:

$$\text{Spiked Recovery Rate} = \frac{\text{the detected amount of ERC (fg)} - \text{the detected amount of un-spiked sample (fg)}}{\text{Theoretical spiked amount (fg)}} \times 100\%$$

The calculation formula can be more detailed as below:

$$\text{Spiked Recovery Rate} = \frac{\text{The detected concentration of ERC (spiked sample)} \times \text{Volume of eluted DNA} - \text{The detected concentration of un-spiked sample (fg/}\mu\text{L)} \times \text{Volume of eluted DNA (}\mu\text{L)}}{\text{Theoretical spiked concentration (fg/}\mu\text{L)} \times \text{Spiked volume (}\mu\text{L)}} \times 100\%$$

NOTE: Calculate the concentration of the test sample (fg/ μ L) based on the standard curve, the Ct value of test sample is only valid for concentration calculation within the assay range of standard curve. When Ct value is outside the range of standard curve, do not use the data to calculate the concentration of test sample.
