

# resDetect™ Human SCF ELISA Kit (Residue Testing) (Enzyme-Linked Immunosorbent Assay)

Catalog Number: RES-A006

Pack Size: 96 tests

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedure

RES06-EN.01

ACTO\*

**INTENDED USE** 

resDetect<sup>TM</sup> Human SCF ELISA Kit (Residue Testing) was developed for the detection and quantitative

determination of human SCF in human serum and cell culture supernates. It is intended for research use

only (RUO).

**BACKGROUND** 

Stem Cell Factor is also known as SCF, kit-ligand, KL, steel factor, KITLG, FPH2, KL-1, Kit l, MGF,

SF, or SHEP7, and is a cytokine that binds to the c-Kit receptor (CD117). SCF can exist both as a

transmembrane protein and a soluble protein. This cytokine plays an important role in hematopoiesis

(formation of blood cells), spermatogenesis, and melanogenesis. The expansion of these cells ex-vivo

(outside the body) would allow advances in bone-marrow transplantation, in which HSCs are transferred

to a patient to re-establish blood formation.

To support the development of CAR-T drugs, ACROBiosystems developed Human SCF ELISA Kit

(Residue Testing) with rigorous methodological validation, which is used evaluation the quality of

CAR-T products in drug development and CMC quality control stages. Besides, this kit can also be used

for the quantitative determination of GMP human SCF Protein (ACROBiosystems, Cat#GMP-SCFH25)

concentrations.

PRINCIPLE OF THE ASSAY

This assay kit is used to measure the levels of human SCF by employing a standard sandwich-ELISA

format. The micro-plate in the kit has been pre-coated with Anti-SCF Antibody. Firstly, add the standard

samples provided in kit and your samples to the plate, incubate and wash the wells. Then add the

Biotin-Anti-SCF Antibody to the plate and form Antibody-antigen-biotinylated antibody complex,

incubate and wash the wells. Next add Streptavidin-HRP to the plate, incubate and wash the wells. At

last, load the substrate into the wells and monitor solution color from blue to yellow. The reaction is

stopped by the addition of a stop solution and the intensity of the absorbance can be measured at 450 nm

and 630 nm. The OD Value reflects the amount of human SCF bound.



# **PRECAUTIONS**

- 1. This kit is for research use only and is not for use in diagnostic or therapeutic applications.
- 2. The kit is suitable for cell supernatant, serum and plasma samples.
- 3. Do not use reagents past their expiration date.
- 4. Do not mix or substitute reagents with those from other kits or other lot number kits.
- 5. If samples generate values higher than the highest standard, dilute the samples with the appropriate calibrator diluent and repeat the assay. If cell supernatant samples need step dilution, except for the final dilution with diluent, other intermediate dilutions can be in cell culture medium.
- 6. Differences in test results can be caused by a variety of factors, including laboratory operator, pipette usage, plate washing technique, reaction time or temperature, and kit storage.
- 7. This kit is designed to remove or reduce some endogenous interference factors in biological samples, and not all possible influencing factors have been removed.

## **MATERIALS PROVIDED**

Table 1. Materials provided

Catalog	Components	Size	Format	Storage		
Catalog	Components	(96 tests)	rormat	Unopened	Opened	
RES006-C01	Pre-coated Anti-SCF Antibody Microplate	1 plate	Solid	2-8°C	2-8°C	
RES006-C02	Human SCF Standard	20 μg	Powder	2-8°C	-70°C	
RES006-C03	Biotin-Anti-SCF Antibody	20 μg	Powder	2-8°C	-70°C	
RES006-C04	Streptavidin-HRP	50 μL	Liquid	2-8°C, avoid light	2-8°C, avoid light	
RES006-C05	10×Washing Buffer	50 mL	Liquid	2-8°C	2-8°C	
RES006-C06	2×Dilution Buffer	50 mL	Liquid	2-8°C	2-8°C	



RES06-EN.01

RES006-C07	Substrate Solution	12 mL	Liquid	2-8°C, avoid light	2-8°C, avoid light
RES006-C08	Stop Solution	7 mL	Liquid	2-8°C	2-8°C

**Note:** It is recommended that Streptavidin-HRP be centrifuged briefly before use to deposit liquid from the tube wall or cap to the bottom of the tube.

## **SRORAGE**

- 1. Unopened kit should be stored at 2°C -8°C upon receiving.
- 2. The opened kit should be stored per Table 1. The shelf life is 30 days from the date of opening.

*Note:* a. Do not use reagents past their expiration date.

b. Find the expiration date on the outside packaging.

## REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or multi-channel micropipettes and pipette tips: need to meet 10  $\mu$ L, 300  $\mu$ L injection requirements;

37°C Incubator:

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Tubes: 1.5mL,10mL;

Timer;

Reagent bottle;

Deionized or distilled water.

# REAGENT PREPARATION

Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, place the sample in an 37°C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.

According to Table 2, prepare the provided lyophilized product into a storage solution with ultrapure water, dissolve at room temperature for 15 to 30 minutes, and mix by gently pipetting, avoiding

3 / 12

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vigorous shaking or vertexing. The reconstituted storage solution should be stored at -70 $^{\circ}$ C. It is recommended that the number of freezing and thawing should not exceed 1 time, the size of the aliquot should not be less than 5  $\mu$ g.

**Note:** Considering inevitable minor quantitation variations between protein batches, it is also reasonable to generate the standard curve with specific lot of proteins used for current production for even better accuracy.

Storage solution ID Reconstituted water Vol. **Components** Size (96 T) concentration. RES006-C02 Human SCF Standard 20 μg  $200 \mu g/mL$  $100 \mu L$ RES006-C03 Biotin-Anti-SCF Antibody  $100 \mu L$ 20 μg  $200 \mu g/mL$ 

Table 2. Preparation method

## RECOMMENDED SAMPLE PREPARATION

# 1. Working Solution Preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

1.2 Preparation of 1×Dilution Buffer:

Dilute 50 mL 2×Dilution Buffer with 1×Washing Buffer to 100 mL.

1.3 Preparation of Biotin-Anti-SCF Antibody working fluid:

Dilute Biotin-Anti-SCF Antibody to 0.4 μg/mL with 1×Dilution Buffer. Please prepare it for one-time use only.

1.4 Preparation of Streptavidin-HRP working fluid:

Dilute Streptavidin-HRP at 1:1000 with 1×Dilution Buffer. The prepared working fluid should avoid light. Please prepare it for one-time use only.

#### 1.5 Sample preparation

If the sample to be tested is the cell supernatant, dilute test sample at 1:10 with 1×Dilution Buffer. The volume ratio of sample to diluent is 1:9.

4 / 12

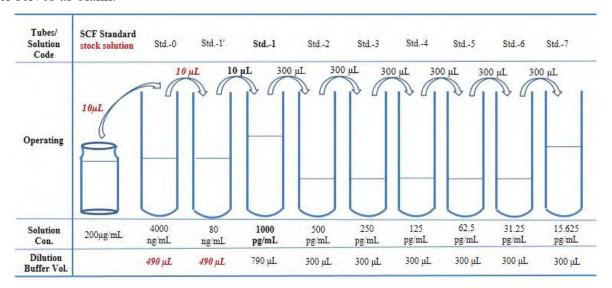
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## 2. Preparation of Standard curve

The concentration of the reconstituted human SCF Calibrator (RES006-C02) is 200  $\mu$ g/mL, prepare (Std.-0) by diluting 10  $\mu$ L the reconstituted human SCF Calibrator into 490  $\mu$ L Sample Dilution Buffer, mix gently well. Then prepare Std.-1' by diluting 10  $\mu$ L Std.-0 into 490  $\mu$ L Sample Dilution Buffer. At last, prepare the highest concentration of standard curve, Std.-1 (1000 pg/mL), by diluting 10  $\mu$ L Std.-1' into 790  $\mu$ L Sample Dilution Buffer. Prepare 1:1 serial dilution for the standard curve as follows: Pipette 300  $\mu$ L of Sample Dilution Buffer into each tube. Make sure to mix well every time. Sample Dilution Buffer serves as blank.



#### 3. Add Samples

Add 100 µL Calibrator and samples to each well. For blank Control wells, please add 100 µL Dilution Buffer.

*Note*: It is recommended to set double holes for samples and standard curves to be tested.

#### 4. Incubation

Seal the plate with microplate sealing film and incubate at 37°C for 1.0 hour.

#### 5. Washing

Remove the remaining solution by aspiration, add 300 µL of 1×Washing Buffer to each well, soak for 10 s, remove any remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it



against paper towels. Repeat the wash step above for three times.

#### 6. Add Biotin-Anti-SCF Antibody

For all wells, add 100 µL Biotin-Anti-SCF Antibody (dilute to 0.4 µg/mL) working solution. Please prepare it for one-time use only.

#### 7. Incubation

Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

#### 8. Washing

Repeat step 5.

#### 9. Add Streptavidin-HRP

For all wells, add 100 µL Streptavidin-HRP (dilute at 1:1000) working solution. Please prepare it for one-time use only, avoid light.

#### 10. Incubation

Seal the plate with microplate sealing film and incubate at room temperature for 30 min.

#### 11. Washing

Repeat step 5.

#### 12. Substrate Reaction

Add 100 µL Substrate Solution to each well. Seal the plate with microplate sealing film and incubate at room temperature for 20 min, avoid light.

#### 13. Termination

Add 50 µL Stop Solution to each well and tap the plate gently to allow thorough mixing.

*Note:* The color in the wells should change from blue to yellow.

#### 14. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 10 minutes.

**Note**: To reduce the background noise, subtract the value read at  $OD_{450nm}$  with the value read at  $OD_{630 nm}$ .

# CALCULATION OF RESULTS

6 / 12

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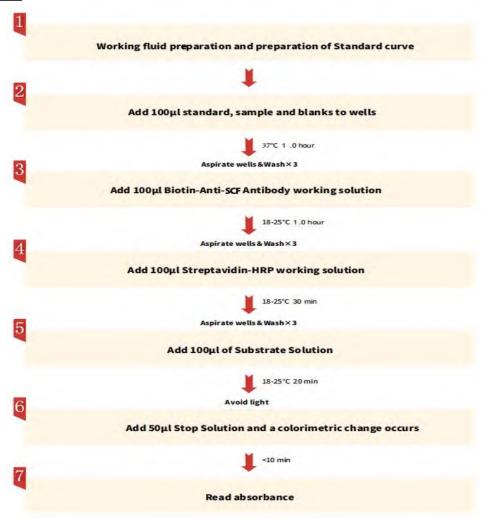
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- 1. Calculate the mean absorbance for each standard, control and sample and subtract average zero standard optical density (OD).
- 2. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.
- 3. Normal range of Standard curve: R<sup>2</sup>≥0.9900.
- 4. Detection range: 15.625 pg/mL-1000 pg/mL. If the OD value of the sample to be tested is higher than 1000 pg/mL, the sample shall be diluted with dilution buffer and assay repeated. If the OD value of the sample to be tested is lower than 15.625 pg/mL, the sample should be reported.

## **QUICK GUILD**

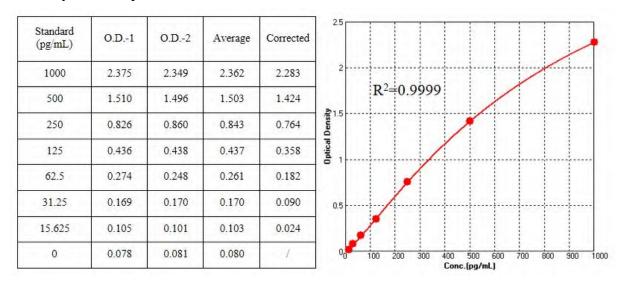


7 / 12



# **TYPICAL DATA**

For each experiment, a standard curve needs to be set for each micro-plate, and the specific OD value may vary depending on different laboratories, testers, or equipments. The following example data is for reference only. The sample concentration was calculated based on the results of the standard curve.



# **SENSITIVITY**

The minimum detectable concentration of SCF is 5.379 pg/mL. The minimum detectable concentration was determined by adding twice standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

### **PRECISION**

1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

	Intra-assay Precision			I	nter-assay Precisio	n
Sample	1	2	3	1	2	3

8 / 12

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n	10	10	10	3	3	3
Mean (pg/mL)	714.833	271.892	33.498	709.324	277.598	31.039
SD	42.639	22.587	2.135	19.359	13.087	2.346
CV (%)	6.0	8.3	6.4	2.7	4.7	7.6

*Note*: The example data is for reference only.

# **RECOVERY**

Five parts of blank T cell culture supernatant were added with different concentrations of human SCF, and the T cell culture supernatant without human SCF was used as background to calculate the recovery rate. The range of the recovery rate is 90.9-101.3%, and the average recovery is 95.3%.

Sample Type	Average % Recovery	Range
T cell culture supernatant (n=5)	95.3	90.9-101.3

# **LINEARITY**

To assess the linearity of the assay, samples spiked with high concentrations of SCF were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture medium	Cell culture medium	T cell culture company tout
		(DMEM)	(1640)	T cell culture supernatant
Average Recovery (%)		96.9	106.3	105.0
1:2	Range (%)	90.0-106.2	101.3-110.6	100.6-109.1
Average Recovery (%)		95.8	108.6	107.9
1:4	Range (%)	85.9-104.4	103.5-111.6	106.6-112.1
1.0	Average Recovery (%)	101.3	113.8	115.4
1:8	Range (%)	92.4-106.6	111.3-116.0	111.3-119.5
1.16	Average Recovery (%)	107.2	114.6	116.6
1:16	Range (%)	97.0-119.1	110.0-119.1	113.4-119.1

*Note*: The example data is for reference only.

9 / 12

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# **SPECIFICITY**

This assay recognizes natural and recombinant human SCF. No cross-reactivity was observed when this kit was used to analyze the following recombinant cytokines.

Human						
IL-2	IL-10	GM-CSF	Anti-CD3			
IL-3	IL-11	G-CSF	Anti-CD28			
IL-4	IL-12B	M-CSF	Anti-CD137			
IL-5	IL-15	IFN-alpha 1	FGF basic			
IL-6	IL-17A	IFN-gamma	Thrombopoietin-TPO			
IL-7	IL-18	TGF-beta 1	L1R			
IL-8	VEGF165	Flt-3 Ligand	BMP-2			
TNF-alpha						

# **INTERFERING SUBSTANCES**

Verify potential matrix effects by adding different levels of DMSO and HSA to the diluted buffer.

Additive	Tolerated concentration
DMSO	20%
HSA	5%

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# **PLATE LAYOUT**

	1	2	3	4	5	6	7	8	9	10	11	12
Α	Std1	Std1	()	()	()		()	()	()	()	()	()
В	Std2	Std2										
c	Std3	Std3	$\left( \begin{array}{c} \cdots \end{array} \right)$									
D	Std4	Std4	$\left( \begin{array}{c} \cdots \end{array} \right)$	$\left( \begin{array}{c} \cdots \end{array} \right)$	()		(iii)	$\left( \begin{array}{c} \cdots \end{array} \right)$	()	$\left( \begin{array}{c} \dots \end{array} \right)$	()	()
E	Std5	Std5	$(\cdots)$	$\left( \begin{array}{c} \cdots \\ \end{array} \right)$	()			()	()	$\left( \begin{array}{c} \cdots \end{array} \right)$	()	()
F	Std6	Std6	$\left( \cdots \right)$	$\left( \begin{array}{c} \cdots \\ \end{array} \right)$	()			()	()	$\left( \begin{array}{c} \cdots \end{array} \right)$	()	()
G	Std7	Std7	()	()	()	;;;;) ;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;	;;;;)	()		()	(	()
Н	Blank	Blank	()	()	()	(iii)	(iii)	()	()	()	()	()

Note: Blank is a Blank Dilution Buffer hole.

# **TROUBLESHOOTING GUIDE**

Problem	Cause	Solution	
Poor standard curve	* Inaccurate pipetting	* Check pipettes	
Lawra CV	* Inaccurate pipetting	* Check pipettes	
Large CV	* Air bubbles in wells	* Remove bubbles in wells	
High background	* Plate is insufficiently washed	* Review the manual for proper wash.	
High background	* Contaminated wash buffer	* Make fresh wash buffer	
Very low readings across	* Incorrect wavelengths	* Check filters/reader	
the plate	* Insufficient development time	* Increase development time	

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Samples are reading too high, but standard curve	* Samples contain cytokine levels above assay range	* Dilute samples and run again
Drift	* Interrupted assay set-up  * Reagents not at room temperature	* Assay set-up should be continuous - have all standards and samples prepared appropriately before commencement of theassay  * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts

Asia and Pacific: