

Streptavidin Coated Plates, Clear, 96-Well

Catalog Number: SP-11 Pack Size: 1 plate / 5 plates

Specifications

Table 1. plate details

Items	Specifications	
Material	Polystyrene	
Color	Clear	
Plate Blocking:	2% BSA Blocking Buffer	
Formulations	Clear, 96-well plates, coated with 100uL of streptavidin tetramer and	
	blocked with 200uL of 2% BSA Blocking Buffer	
Detection Method	Colorimetric	
Capacity	~5pmol biotin/well	
CV% of plates/wells	< 10%	
Туре	Detection Plate, Immunoassay, ELISA	

Shipping and Storage

Upon receipt store plates at 4°C in unopened pouches. Once opened, place unused plates in a resealable bag with desiccant and store at 4°C. This plate is supplied and shipped with blue ice.

Product description

The Streptavidin Coated Plates, Clear, 96-Well is pre-coated with Streptavidin tetramer protein and blocked with BSA, it is a ready-to-use polystyrene plate, which can be used for binding biotinylated proteins and antibodies, or probes for ELISA and other target-specific assays. The recombinant Streptavidin is tetramer protein expressed in E. coli designed for immobilization applications.

<u>Applications</u>

This Streptavidin Coated Plate is intended for Immunoassay and ELISA.

IMPORTANT: Please carefully read this manual before performing your experiment.

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Assay Principles

Streptavidin (SA) has an extraordinarily high affinity for biotin with a dissociation constant (Kd) on the order of 10⁻¹⁴ mol/L, the Biotinylated molecules can bind to the SA irreversibly. Streptavidin has an isoelectric point of 5 to 6, resulting in low nonspecific interactions. The Streptavidin Coated Plates we provide are easy to use and widely available for applications.

Example ELISA Procedure

Materials and Reagents Preparation

Before starting the ELISA Assay, we should prepare the all reagents and materials required in the experiment. You can prepare these regents by following operations, we also provide the matching reagent kit (Cat. No. SP-11).

Wash Buffer: PBS or TBS with 0.05% (v/v) Tween-20 (usually at pH7.4), 500 mL is sufficient for 96 tests. The pH of Buffer system can be adjust according to your experiment.

Dilution Buffer: Wash Buffer with 0.5% (w/v) bovine serum albumin (BSA) (i.e. Jackson, Catalog#. 001-000-162), 50 mL is sufficient for 96 tests.

Substrate Dilution Buffer: 50 mM disodium hydrogen phosphate (Na_2HPO_4) and 25 mM citric acid, adjust pH to 5.5 with 1 M Sodium hydroxide (NaOH), 25 mL is sufficient for 96 tests.

Substrate Stock Solution: 20 mg/mL TMB (*Sigma-Aldrich, Catalog # 860336*) in Dimethyl sulfoxide (*Sigma-Aldrich, Catalog # D8418*), 1 mL is sufficient for 96 tests. **Protect from light**.

TMB Substrate Working Solution

For **each plate** dilute 125 μ L substrate stock solution in 25 mL substrate dilution buffer and add 20 μ L 5% H₂O₂ (pipette 10 μ L 30% H₂O₂ into 50 μ L distilled water), mix well.

Notes:

- 1) The TMB Substrate Working Solution should be freshly prepared and used within 15 minutes.
- 2) If you choose to use other commercially available ready-to-use TMB substrate solutions, you should follow the manufacturer's instruction.

Stop Solution: 1 M sulfuric acid (aqueous), 6 mL is sufficient for 96 tests.

Microplate sealing film (Sigma-Aldrich, Catalog # Z724742)

Pipettes and pipette tips

UV/Vis microplate spectrophotometer (absorbance 450 nm, correction wavelength set to 630 nm).

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Recommended Protocol

1. Preparation

Reconstitute and store all reagents as recommended.

2. Washing

Add 300 μ L of Wash buffer to each well, gently tap the plate for 1 minute, remove any remaining Wash Buffer by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

3. Add biotinylated protein or antibodies

- 1) Dilute Biotinylated protein or antibodies to a concentration you want (usually $1^{10} \mu g/mL$) with Dilution Buffer to make Biotinylated molecule working solution.
- 2) Add 100 μ L Biotinylated molecule to each well and incubate at 37 $^{\circ}$ C or RT for 1 hour.
- 3) For the "Blank" wells, please add 10 μ L Biotinylated molecule working solution.
- 4) For Non specific of the sample wells, please add 100 μ L Dilution Buffer.

4. Washing

Remove the remaining solution by aspiration, add 300 μ L of Wash buffer to each well, gently tap the plate for 1 minute, remove any remaining Wash Buffer by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above **for three times**.

5. Add Samples

- 1) Make series dilution of the samples as appropriate with Dilution Buffer.
- 2) Add 100 µL of the serial dilution of sample to each well, incubate at 37 °C or RT for 1 hour.

6. Washing

Repeat step 4.

7. Add primary antibody

- 1) Dilute primary antibody to an appropriate concentration with Dilution Buffer.
- 2) For all wells, add 100 μ L of diluted primary antibody, and incubate at 37°C or RT for 1 hour.

8. Washing

Repeat step 4.

9. Add enzyme-labeled secondary antibody

- 1) Dilute secondary antibody to an appropriate concentration with Dilution Buffer.
- 2) For all wells, add 100 μ L of diluted secondary antibody, and incubate at 37 $^{\circ}$ C or RT for 1 hour, avoid light.

10. Washing

Repeat step 4.

11. TMB Substrate Reaction

Add 200 μ L TMB Substrate Working Solution to each well. Seal the plate with microplate sealing film and incubate at 37 $^{\circ}$ C or RT for 20 minutes, avoid light.

12. Termination

Add 50 μ L **Stop Solution** to each well, and tap the plate gently for 3 minutes to allow thorough mixing

Note: the color in the wells should change from blue to yellow.

13. Data Recording

Read the absorbance at 450 nm using UV/Vis microplate spectrophotometer.

Note: the plate may be read at 600 nm without adding 1 M sulfuric acid, but the Signal-to-Background ratio

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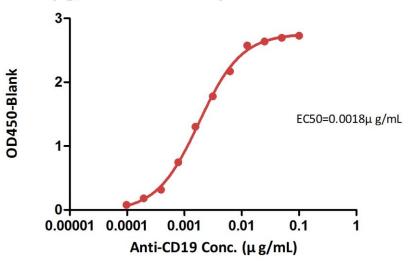
may be reduced.

Example Data

1. Binding Assay between CD19 and Anti-CD19 antibody on SA Plate

Immobilized Biotinylated Human CD19 (20-291), His,Avitag (Cat. No. CD9-H82E9) at 1 μ g/mL (100 μ L/well) on Streptavidin Coated Plates, Clear, 96-Well (Cat. No. SP-11), can bind Anti-FMC63 antibody with a linear range of 0.1-3 ng/mL (QC tested).

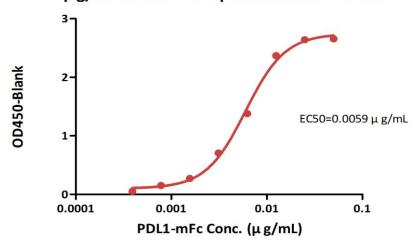
5 μg/mL SA Coated onto Plate per well 1 μg/mL Biotin-CD19 captured to the SA-Plate



2. Binding Assay between PD1 and PDL1 on SA Plate

Immobilized Biotinylated Human PD-1, Fc, Avi tag, His Tag (Cat. No. PD1-H82F4) at 1 μ g/mL (100 μ L/well) on Streptavidin Coated Plates, Clear, 96-Well (Cat. No. SP-11), can bind Human PD-L1, mFc Tag with a linear range of 0.4-6.25 ng/mL (QC tested).

 $5 \mu g/mL$ SA Coated onto Plate per well $1 \mu g/mL$ Biotin-PD1 captured to the SA-Plate



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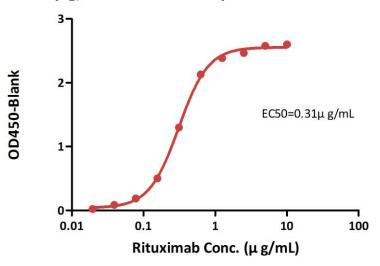
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3. Binding Assay between CD20 and Rituximab on SA Plate

Immobilized Biotinylated Human CD20 Full Length, His, Avi tag (HEK293) (Cat. No. CD0-H82E5) at 1 μ g/mL (100 μ L/well) on Streptavidin Coated Plates, Clear, 96-Well (Cat. No. SP-11), can bind Rituximab with a linear range of 78-625 ng/mL (QC tested).

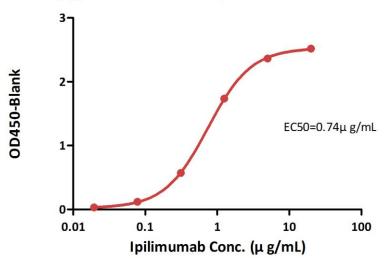
 $5 \mu g/mL$ SA Coated onto Plate per well $1 \mu g/mL$ Biotin-CD20 captured to the SA-Plate



4. Binding Assay between FcRn and Ipilimumab on SA Plate

Immobilized Biotinylated Human FCGRT&B2M Heterodimer Protein, His, Avi tag (Cat. No. FCM-H82W7) at 1 μ g/mL (100 μ L/well) on Streptavidin Coated Plates, Clear, 96-Well (Cat. No. SP-11), can bind Ipilimumab with a linear range of 78-1250 ng/mL (QC tested).

$5 \mu g/mL$ SA Coated onto Plate per well $1 \mu g/mL$ Biotin-FcRn captured to the SA-Plate



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TROUBLESHOOTING GUIDE

Problem	Possible Cause	Solutions
	Incorrect storage of plate	The plate should be store plates at 4°C, once you open the package, get the amount you need and keep the rest airtight.
Signal of positive control is weak or abnormal	Detection Antibody is outdated or no prepared the working solution immediately before use	The working solution should be prepared immediately before use and should not be stored.
	Errors in instrument settings	♦ Please check instrument setting.
	SubstrateStockSolutionisoutdated;Incubationtemperatureisincorrect;Incubation time is not sufficient;Repeated freeze-thaw cycles;	 Make sure the Substrate Stock Solution is working. Use proper incubation time and temperature.
	Pipetting errors	♦ Make sure that the pipette is calibrated and working properly.
High background	Serum samples	If you want test serum samples, the BSA Blocking plate is not suitable for this purpose. We specially developed the Streptavidin Coated Plates, Clear, 96-Well (For Serological Testing) (Cat.No.SP-13) for serological testing.
	Sample solvent contains inhibiting factors	 Run a negative control assay with the solvent alone. → Maintain DMSO level at <1%. Increase protein incubation time.
	Contamination	Make sure buffers and samples are prepared, used and stored correctly.
	The TMB Substrate Working Solution is not fresh	TMB Substrate Working Solution must be used within 15 minutes after preparation.
Colorimetric signal is erratic	Inconsistent pipetting or dilution methods	 → Make sure pipettors are functioning properly and use a multichannel pipettor if possible. → Use master mixes to minimize errors. → Run duplicates for all tests.
	TMB Substrate Working Solution is not completely mixed with the reaction solution	Make sure that TMB Substrate Working Solution is adequately mixed with the reaction solution.
	Bubbles in the wells	→ Tap plate gently to disperse bubbles.
	Signal is too high	 The concentration of the samples should be adjusted to achieve optimal reading. Decrease colorimetric HRP substrate incubation time.
Inadequate color development	Incomplete removal of residual buffers during previous steps	♦ Wells should appear dry after aspiration.
	Problems with conjugate or color reagents	Color should appear immediately after the reagent is added. Make sure no contamination or residual buffers in the wells before you start the color development process.

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